

Better Training for Safer Food BTSF

FISH DISEASES OF CONCERN UNDER EU LEGISLATION

GIUSEPPE BOVO

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INTRODUCTION-1

According to annex IV, part II, of 2006/88/EC directive the following diseases are regarded of major concern:

EXOTIC

Epizootic ematopoietic necrosis (EHN)

Epizootic ulcerative syndrome (EUS)

NON EXOTIC

Viral haemorrhagic septicaemia (VHS)

Infectious haematopoietic necrosis (IHN)

Koi herpesvirus disease (KHVD)

Infectious salmon anemia (ISA)





INTRODUCTION -2

In addition to the previous notifiable diseases the following diseases may be considered at national level and MS may require additional guarantees (Dec. 2004/453)

Bacterial kidney disease (BKD)
Spring viraemia of carps(SVC)
Gyrodactylus salaris (Gs)
Infectious pancreatic necrosis (IPN)





EPIZOOTIC ULCERATIVE SYNDROME

- EUS -





DEFINITION AND HISTORY

EUS IS A SEASONAL EPIZOOTIC PATHOLOGICAL CONDITION AFFECTING SEVERAL FARMED AND WILD FISH SPECIES BOTH IN FRESH AND BRACKISH WATER ENVIRONMENT





AETIOLOGY

EUS HAS A COMPLEX AETIOLOGY

THE INVASIVE OOMYCETE

APHANOMICES INVADANS, IS
CONSIDERED THE RESPONSIBLE AGENT
FOR INDUCING THE TYPICAL LESIONS:
ULCERS AND GRANULOMAS

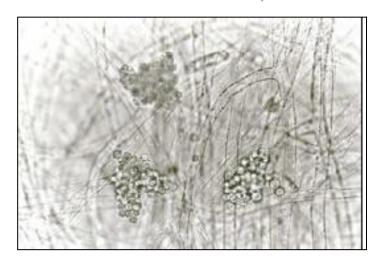
THE TAXONOMY OF THE AGENT, A NON SEPTATE FUNGUS-LIKE IS NOT DEFINITIVE (Diatoms, brown algae)

CONCURRENT INFECTIONS WITH BACTERIA AND PARASITES MAY BE OFTEN DETECTED

Musculature smear showing typical Aphanomyces sporangia (library .enaca.org.)



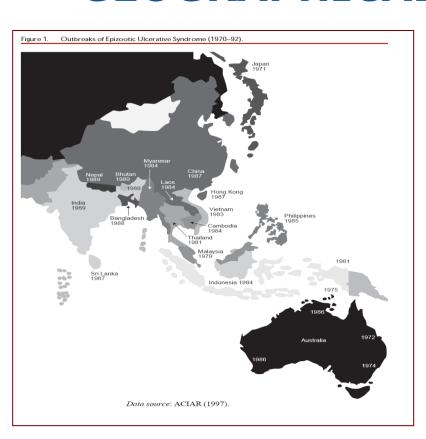
Daniel J. Fairweather, BSc.







GEOGRAPHICAL DISTRIBUTION



ENDEMIC:

First detection in Japan (1971), South east Asia, South Asia, Australia.

RECENT REPORTS:

Canada (2011) USA

Central Africa (Botswana, Namibia, Zambia)

NOAA Fisheries





SUSCEPTIBLE HOSTS

TARGET SPECIES: > 76 SPECIES AFFECTED

- Black bullhead
- Grey mullet & other mullets
- Well catfish

RESISTENT SPECIES:

- Common carp,
- Nile Tilapia,





ORIGIN AND TRANSMISSION

WATER CONTAMINATED WITH THE ZOOSPORES IS THE MAIN INFECTION SOURCE

SECONDARY ZOOSPORES MAY INCIST AND SURVIVE IN THE ENVIRONMENT WAITING A SUSCEPTIBLE HOST

ORNAMENTAL FISH FROM ENDEMIC AREAS (i.e. Gourami spp.) MAY PLAY AN IMPORTANT ROLE IN SPREADING THE INFECTION





MORTALITY AND MORBIDITY

SEVERE EPIZOOTICS OCCUR DURING COLD SEASONS (18-22 °C)

MORBIDITY AND MORTALITY OFTEN > 50%

LOW TEMPERATURE AND HEAVY RAINFALL IMPROVES SPORULATION CAUSING AN INCREASE OF OUTBREAKS

OUTBREAKS IN FARMS OFTEN SECONDARY TO EPIZOOTICS OCCURRING IN WILD POPULATION





CLINICAL SIGNS

LOSS OF APPETITE

DARKENING OF THE SKIN

SWIMMING NEAR THE WATER SURFACE

RED SPOTS AND SKIN ULCERS

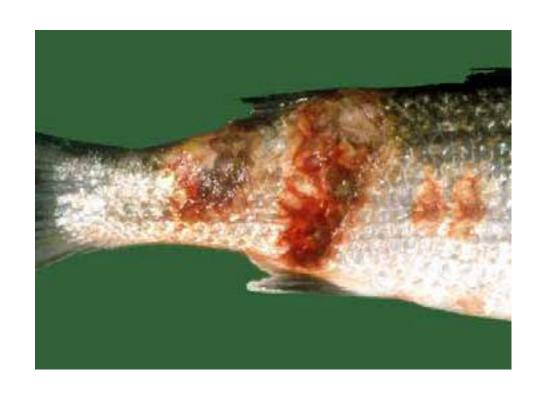




GROSS PATHOLOGY

RED OR GREY LARGE ULCERS IN LATER STAGES

EROSION OF THE MUSCULATURE, AND SOFT AND HARD TISSUES OF THE CRANIAL AREA WITH BRAIN EXPOSURE



J. D. Humphrey and M. Pearce, Fishnote 2004





MICROSCOPIC PATHOLOGY

ERYTHEMATOUS DERMATITIS

PRESENCE OF APHANOMICES INVADANS HYPHAE

GROWING IN THE SKELETAL MUSCULATURE

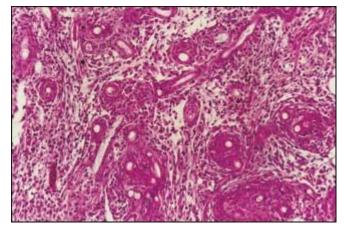
NECROTISING GRANULOMAS

SECONDARY MYCOTIC, BACTERIAL AND

PARASITIC INVASION

M.G. BONDAD-REANTASO

Typical severe mycotic granulomas from muscle section of EUS fish (H & E)







The diagnosis of EUS is based on clinical signs and confirmed by histopathology

SELECT 10 FISH WITH TYPICAL LESION IN EARLY STAGES KEEP THE SAMPLES AT 4 °C FOR 1-2 DAYS OR FIX THEM IN BUFFERED FORMALIN DIAGNOSIS IS CONFIRMED BY:

OBSERVATION OF THE HYPHAE IN FRESH TISSUES ISOLATION ON PEPTONE/GLUCOSE AGAR

DETECTION BY HISTOLOGY DETECTION BY PCR METHODS

M.G. BONDAD-REANTASO Mycotic granulomas showing fungal hyphae (stained black) using Grocotts stain





CONTROL AND PREVENTION

NO VACCINES AVAILABLE

NO CHEMICAL TREATMENT AVAILABLE

TREATMENT OF INFECTED FARMS WITH LIME OR HIDRATED LIME MAY REDUCE THE IMPACT OF THE DISEASE.

THE EXOTIC STATUS IN EU?





EPIZOOTIC HAEMATOPOIETIC NECROSIS - EHN -





DEFINITION AND HISTORY

EHN IS AN EXOTIC SISTEMIC VIRAL DISEASE AFFETCING WILD PERCH POPULATIONS (PERCA FLUVIATILIS) IN AUSTRALIA (LANGDON ET AL 1986)

FOLLOWING THE FIRST REPORT IN PERCH THE DISEASE HAS BEEN OBSERVED IN FARMED RAINBOW TROUT (LANGDON ET AL., 1986)





AETIOLOGY

The causative agent of EHN is a member of the family IRIDOVIRIDAE, genus Ranavirus, FROG VIRUS 3 (FV3) being the type species:

MORFOLOGY : Icosahedral

SIZE : 150-180 nm

BIOCHEMISTRY : 1 ds DNA,

Robust serological and genomic homologies are shown with different ranaviruses causing similar diseases in different species

•	Japanese seabass (Lates calcarifer)	LMBV
•	Pikeperch (Stizostedion lucioperca)	PPIV
•	Short finned eel (Anguilla australis)	SERV
•	European catfish (Silurus glanis)	ESV
•	Blackbullhead (Ameiurus melas)	ECV
•	B.cleaner wrasse (Labroides dimidiatus)	DFV
•	Guppy (Poecilia reticulata)	GV6





SUSCEPTIBLE HOSTS

REDFIN PERCH (PERCA FLUVIATILIS) IN WILD POPULATION

RAINBOW TROUT (ONCORHYNCHUS MYKISS)

IN FARMS CONNECTED WITH RIVERS SUFFERING EHN OUTBREAKS

THE DISEASE MAY AFFECT FISH AT ALL LIFE CLASSES BUT IS MORE FREQUENT AMONG JUVENILES

SEVERAL DIFFERENT SPECIES MAY DEVELOP THE DISEASE WHEN EXPERIMENTALLY INFECTED BY BATH



Perca fluviatilis picture Kohout J



O. mykiss picture (Onmyk_f0.jpg) by McDowall, R.M.





GEOGRAPHICAL DISTRIBUTION

THE DISEASE HAS ONLY BEEN REPORTED FROM SOME REGIONS IN THE AUSTRALIAN CONTINENT





ORIGIN AND TRANSMISSION

EHN IS HORIZZONTALLY TRANSMITTED; NO DATA ON VERTICAL TRANSMISSION ARE AVAILABLE

NEW OUTBREAKS IN FARMED FISH SEEMS TO BE CONNECTED WITH NATURAL EPIZOOTIC OCCURRING IN THE WILD POPULATION

THE VIRUS MAY BE TRANSPORTED BY NETS, BOATS, AND OTHER EQUIPMENTS

BAIT FISH ARE CONSIDERED AS A POTENTIAL RISK

PISCIVOUROUS BIRDS ARE CONSIDERED AS A POTENTIAL RISK (WITTINGTON, 1996)

AMPHIBIA ARE REGARDED AS NATURAL RESERVOIR





MORTALITY AND MORBIDITY

TROUT

OUTBREAKS OCCUR AT 11-20° C LOW DIRECT ECONOMIC LOSSES, MAINLY RELATED TO OVERCROWDING, BAD WATER QUALITY AND INTERCURRENT DISEASES.

REDFIN PERCH

OUTBREAKS OCCUR AT HIGH TEMPERATURE
IN SUMMER BOTH MORBIDITY AND MORTALITY BECOME
IMPORTANT LEADING TO A SIGNIFICANT LOSS OR REDUCTION
OF WILD POPULATION, PARTICULARLY WHEN LARGE NUMBER
OF JUVENILES ARE PRODUCED







Mass mortality of redfin perch. Note the small size of individuals affected and swollen stomach of fish at the centre of the photograph

Source: anonymou

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CLINICAL SIGNS

According to the OIE ref. Lab there are no specific clinical signs. Affected fish may show:

- LOSS OF EQUILIBRIUM
- FLARED OPERCULA
- DARKENING OF THE SKIN





GROSS AND MICROSCOPIC PATHOLOGY

EHNV is an endotheliotropic agent causing extensive haemorrhagies, oedema and reduction of peripheral circule. A small proportion of fish may have:

KIDNEY, LIVER AND SPLEEN ENLARGMENT

The most prevalent lesions are represented by focal or extensive necrosis interesting:

HAEMATOPOIETIC KIDNEY SPLEEN





SAMPLING AND DIAGNOSIS

Select 10 specimens showing clear signs referrred to EHN and submitt them, in refrigerated box, to the laboratory:

- VIRUS ISOLATION AND IDENTIFICATION
 Liver
 Anterior kidney
 Spleen
- VIRUS IDENTIFICATION BY ELISA
- PCR + REA (according to the OIE manual) to differentiate from different ranaviruses





CONTROL AND PREVENTION

Because no commercial vaccine is available the impact of the disease may be reduced through

REDUCTION OF THE STOCKING DENSITY GOOD QUALITY WATER SUPPLY





INFECTIOUS SALMON ANEMIA

- ISA -





DEFINITION AND HISTORY

ISA IS A SISTEMIC LETHAL VIRAL DISEASE, CAPABLE TO INDUCE SEVERE ANEMIA AND HEAVY LOSSES IN ATLANTIC SALMON (Salmo salar) DURING THE SEA-FARMING CYCLE.





AETIOLOGY

The causative agent of ISA is an RNA virus belonging to the family ORTOMYXOVIRIDAE, genus ISAVIRUS:

MORFOLOGY: PLEOMORPHIC PARTICLES

DIZE : 100-130 nm

BIOCHEMISTRY: 8 SS RNA, > 10 STRUCTURAL PROTEINS

BIOPHISIC: INACTIVATION 30 min a 56° C

INACTIVATION Cl₂ 100 ppm 15 min

INACTIVATION O₃ 8 mg/l 4 min

INACTIVATION pH 4 e pH 12 x 24 h.





SUSCEPTIBLE HOSTS

ATLANTIC SALMON (SALMO salar) IS THE MAIN TARGET SPECIES. MOST OUTBREAKS HAVE BEEN REPORTED FROM MARINE ENVIRONMENT

THE DISEASE HAS BEEN ALSO REPORTED FROM SILVER SALMON (Oncorhynchus kisutch) IN CHILE

RAINBOW TROUT (O. mikiss) IS CONSIDERED A POTENTIAL CARRIER (detection of the virus in Ireland)

ISAV HAS BEEN FURTHERMORE ISOLATED FROM HEALTHY ATLANTIC COD (Gadus morhua) AND POLLOCK (Pollachius virens) REARED IN THE SAME CAGE WITH ATLANTIC SALMON SHOWING ISA CLINICAL SIGNS



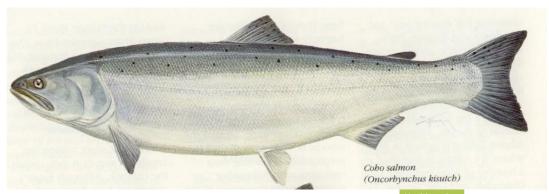




Salmo salar - Sasal_u5.jpg) by McDowall, R.M.



O. mykiss picture (Onmyk_f0.jpg) by <u>McDowall, R.M.</u>



Oncorhynchus kisutch pond.dnr.cornell.edu

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GEOGRAPHICAL DISTRIBUTION

ISA was first observed in 1984 in a hatchery located along the west norvegian coast. In the following years the disease spread the southern coast.

To date ISA has been diagnosed in the following areas :

GEOGRAPHICAL AREA	YEAR	REFERENCES
NORWAY	1984	
CANADA (New Brunswick)*	1996	Bouchard et al.,
SCOTLAND	1998	Rodger et al.
CANADA(Nova Scotia)	2000	_
FAROER	2000	report OIE
USA (Maine)	2001	-
CHILÈ	2001	Kibenge et al.,

^{*} Preliminarly reported as HKS





ORIGIN AND TRANSMISSION

ISA IS HORIZZONTALLY TRANSMITTED

CONTAMINATED WATER

WILD FISH

SHARING STAFF

WELL BOATS

SEA LICE(*Lepeophtheirus salmonis*)

VERTICAL TRANSMISSION HAS BEEN PROPOSED BY SOME AUTHORS (Nylund et al., 2007)

VIRUS ENTRY:

MOST LIKELY THROUGH GILLS. ENTRY THROUGH THE INTESTINE HAS BEEN PROPOSED

VIRUS SHEDDING: THROUGH NATURAL XCRETIONS/SECRETIONS





CLINICAL SIGNS

LETHARGY WITH FISH KEEPING CLOSE TO THE NET PEN PALE GILLS EXOPHTHALMIA DISTENDED ABDOMEN PETECHIAE ON THE SKIN HAEMORRHAGIES IN THE ANTERIOR EYE CHAMBER DURING THE LAST PHASE FISH REMAIN ON THE BOTTOM OF THE CAGE NO FEED IN THE DIGESTIVE TRACT





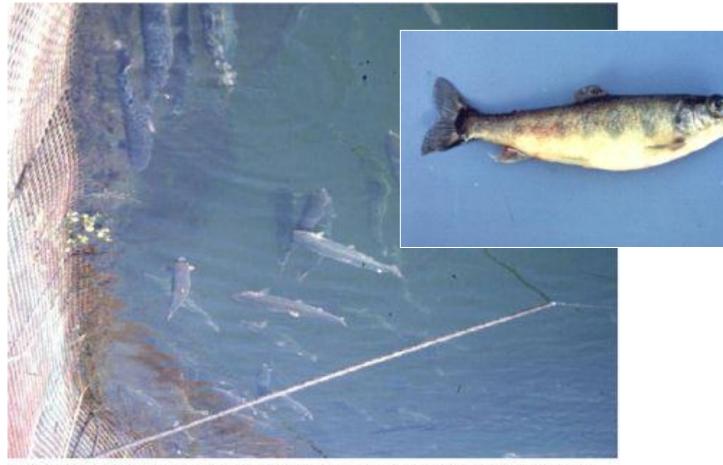


Figure 3. Moribund (sick) Atlantic salmon in an ocean netpen floating on their sides at water surface. Fish are clinically ill form infection by Infectious Salmon Anemia virus. Image from M. Opitz University of Maine.

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GROSS PATHOLOGY

YELLOWISH/ REDDISH LIQUID IN CARDIAC AND PERITONEAL CAVITY

LIVER, KIDNEY AND SPLEN AND INTESTINE CONGESTION

FOCAL HAEMORRHAGIES AND NCROSIS IN THE LIVER

DARK RED LIVER

GILL AND CARDIAC ANEMIA

HAEMORRHAGIES IN THE MUSCULATURE AND PERIVISCERAL FAT

OEDEMA OF THE SWIM BLADDER (CHRONIC STAGE)

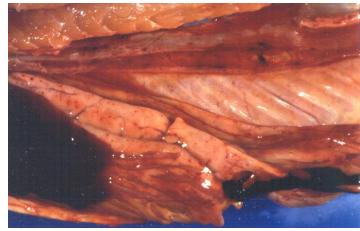














MICROSCOPYC PATHOLOGY

MULTIFOCAL/EXTENSIVE HAEMORRHAGIES WITH NECROSIS FOCI IN THE LIVER .

ERYTHROCYTES ACCUMULATION IN THE SPLEEN STROMA, IN KIDNEY GLOMERULI AND IN BLOOD VESSELS OF THE INTESTINAL LAMINA PROPRIA

SLIGHT MULTIFOCAL/EXTENSIVE INTERSTITIAL HAEMORRHAGIES WITH TUBULAR NECROSIS.

ERYTROPHAGOCYTOSIS IN THE SPLEEN AND HAEMORRHAGIES IN LIVER AND KIDNEY





CLINICAL CHEMISTRY

DEGENERATION AND VACUOLATION OF RED BLOOD CELLS

LEUCOCITES REDUCTION

HEMATOCRIT < 10 (FINAL STAGE)

HAEMATOCRIT 25-30 (INTERMEDIATE STAGE)

LIVER ENZIMES >>





MORBIDITY AND MORTALITY

SIGNIFICANT DIFFERENCES AMONG CAGES AT FARM LEVEL AND BETWEEN FARMS

DAILY MORTALITY 0.5-1%

PEAKS IN EARLY SUMMER AND WINTER

IN SERIOUS OUTBREAKS CUMULATIVE MORTALITY > 90%

STRESS (HANDLING OF FISH) MAY INITIATE DISEASE OUTBREAK





DIAGNOSIS (Dec. UE 2003/1831)

ISA suspicion is confirmed when A or B or C criteria are satisfied

A) CLINICAL SIGNS CONSISTENT WITH ISA + DETECTION OF ISAV:

isolation in cell culture and identification by PCR in tissues (ie BY IF)

- B) VIRUS ISOLATION AND IDENTIFICATION OF ISAV FROM 2 SAMPLES COLLECTED ON SEPARATE OCCASIONS
- C) VIRUS ISOLATION AND IDENTIFICATION FROM 1 SAMPLE + POSITIVE PCR OR IFAT FROM TISSUES

DIFFERENTIAL DIAGNOSIS

Different anemic and haemorrhagic pathological conditions Winter ulcers and septicaemia caused by Moritella viscosa





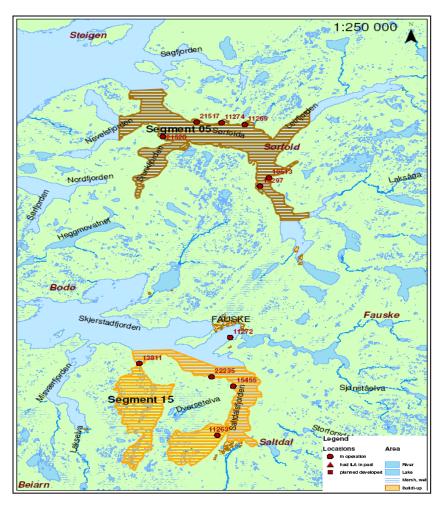
CONTROL AND PREVENTION

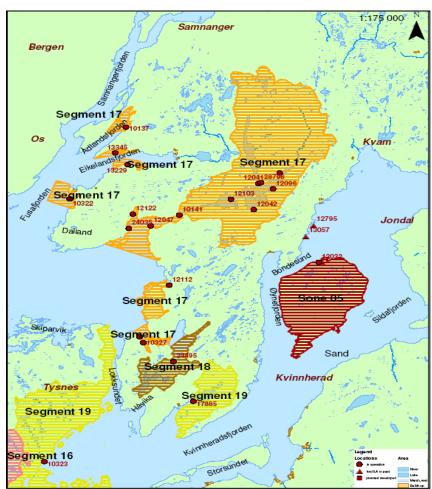
Vaccination has been used in North America and Faroes Islands with no complete protection and risk to establish carriers. The prevention and control of ISA may be obtained according to the following measures:

- CATEGORIZATION OF COMPARTMENTS
- STRICT SURVEY OF CONTAINEMENT ZONE
- PROHIBITION OF VACCINATION
- TRANSPORT AND SLAUGHTERHOUSE REGULATION
- DISINFECTION OF TRUCKS AND WELL BOATS
- TRADE RESTRICTION FROM INFECTED AREAS
- EGG DISINFECTION
- CONTROL OF VECTORS (Lepeophtheirus salmonis)











VIRAL HAEMORRHAGIC SEPTICAEMIA

- VHS -





DEFINITION AND HISTORY

VHS is a sistemic disease characterized by haemorrhagic picture affecting several species both from freshwater and marine environment.

The disease has been observed since 1938 in Rainbow trout (O. Mykiss) which is the major affected species and described with different sinonims

All the VHS isolates react with the MAb IP5B11





The causative agent is an RNA virus belonging to the *Rhabdoviridae* family, genus *Novirabdovirus*. 4 main genotypes described

ТҮРЕ	PREVALENT HOST TYPE AND LOCATION				
I-a	Farmed rainbow trout and a few other freshwater fish in continental Europe ^[10]				
I-b	Marine fish of the Baltic Sea, Skagerrak, Kattegat, North Sea, Japan ^[1]				
I-c	Farmed rainbow trout Denmark				
I-d	Farmed rainbow trout in Norway, Finland, Gulf of Bothnia				
I-e	Rainbow trout in Georgia, farmed and wild turbot in the Black Sea ^[11]				
II	Marine fish of the <u>Baltic Sea</u>				
Ш	Marine fish of the British Isles and northern France, farmed turbot in the UK and Ireland, and Greenland halibut (<i>Reinhardtius hippoglossoides</i>) in Greenland. Rainbow trout in Norway (marine environment				
IV-a	Marine fish of the Northwest Pacific (North America), North American north Atlantic coast, [13] Japan, and Korea [11][14]				
IV-b	Freshwater fish in North American Great Lakes region ^[14]				

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Origin: OIE diagnostic manual



SUSCEPTIBLE HOSTS

In the last 20 years the causative agent has been isolated from > 80 species. In farmed condition the target species is RT but the disease has been observed in different species. According to Directive 2006/88 susceptible species are:

FRESHWATER

Brown trout Salmo trutta fario

Northern pike Esox lucius (Meyer, 1985)

Grayling Thymallus thymallus (Meyer and Whali, 1988)

Whitefish Coregonus sp. (Meyer et al., 1986)

MARINE

Turbot Scopthalmus maxim.(Schlotfeldt et al., 1991)

European cod Gadus morhua

Pacific cod GAdus macrocephalus

Pacific herring Clupea pallasii

Spratt Sprattus sprattus

Pacific salmon Oncorhynchus spp Haddock Gdus aeglefinus

Rocking Onos mustelus

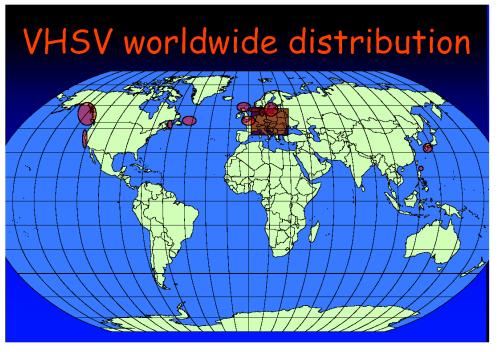




GEOGRAPHIC DISTRIBUTION

VHS in trout has been only observed in Europe, representing one of the most serious disease affecting salmonid aquaculture. The disease is still present in:

AUSTRIA
BELGIUM
BULGARIA
CZECH REPUBLIC
FINLAND
GERMANY
ITALY
NETHERLANDS
POLAND
SLOVAKIA
SLOVENIA
SWITZERLAND



Courtesy prof. N.J. Olesen Fish Diseases EU Ref. Lab. (Aarhus –DK)



June 06

Lake St. Clair

Date	Location	State or Province	Commissi Primary species (Other species)	Estimated	Comments		
2003-05	Lake St. Clair	MI	Muskellunge	4 of 27	Samples submitted over several years		
Summer 05	Bay of Quinte/Lake Ontario	Ontario	Freshwater Drum (Muskellunge Round Goby)	Several hundred tons	Very large natural mortality		
May 06	Sandusky Bay/Lake Erie	ОН	Freshwater Drum	Very large mortality	"Windrows" of fish on beach		
May 06	St. Lawrence River	NY	Round Goby (Muskellunge)	Large die off	River origin		
May 06	Lake Erie	Dry S		Large die off	Fish dying in commercial traps		
May 06	Lake Ontario	Huron Ontario Toro	St. Lawrence River	Mortality event	Acute mortality - no external signs		
May 06	Lake Erie	Lake St. Clair	New York Buffalo	Mortality in wild	Samples from area of traps and mortality		

Pennsylvania

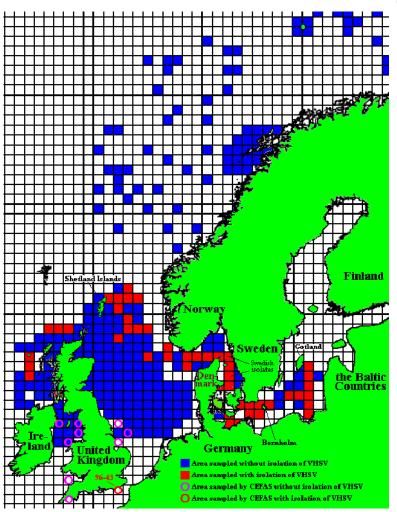
(Yellow perch)

Large mortality

Cleveland

Ohio





VHS INFECTION IN WILD MARINE FISH

Epidemiological investigations carried out by CEFAS (UK) And by EU- RL Aarhus (DK)

Data from EU Fish Diseases Reference Laboratory (Aarhus –DK).

Consumers



ORIGIN AND TRANSMISSION

VIRUS ENTRY : GILLS ARE THE MAJOR PORTAL

VIRUS REPLICATION: FIRST IN THE EPITELIAL CELLS OF THE GILLS

FOLLOWING THE TISSUE INVASIONS

EPLICATION TAKE PLACE IN THE ENDOTHELIA

VIRUS SHEDDING: URINE, OVARIAN FLUID AND SKIN

NOTE . EGGS MAY TRANSMIT THE INFECTION TO THE OFFSPRING BY EXTERNAL CONTAMINATION .





SIGNS AND PATHOLOGICAL CHANGES

The infection is characterized by tree major phases

ACUTE

Lethargy, mono-bilatera exopthalmus, hepatic congestion, haemorrhagies, high mortality

SUB-ACUTE - CHRONIC

Iperpigmentation exopthalmus, liver and kidney anemia anemia epatica e renale, moderate mortality

NERVOUS

Abnormal swimming, kidney anemia, low mortality





EXTERNAL SIGNS

Affected fish are lethargic and remain along the wall of the concrete tanks or, in earthern ponds, near the surface of the water lying on the crevices of the banks. The main signs:

DARKENING OF THE SKIN
SEVERE GILL ANEMIA
EXOPTHALMUS MONO-BILATERAL
PERIORBITAL HAEMORRHAGIES
ABDOMEN DISTENSION





European Commission









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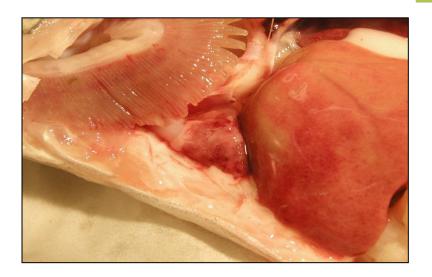
GROSS SIGNS

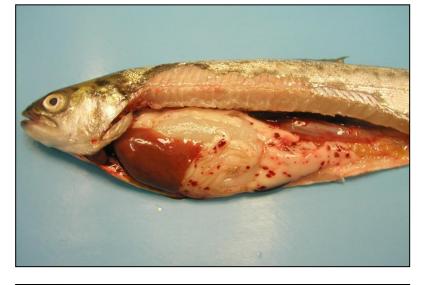
Main internal signs are characterized by the presence of ascitic fluid and a diffuse haemorrhagic picture intersting:

SKELETAL AND CARDIAC MUSCULATURE
LIVER
KIDNEY
SWIMBLADDER
PERIVISCERAL FAT
PERITONEUM

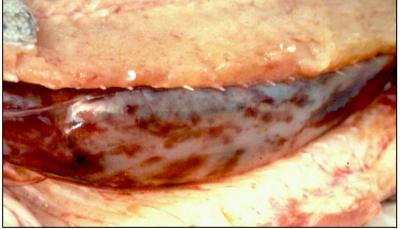














DIFFERENTIAL DIAGNOSIS

DISEASE

LESIONS

TEST

Gas bubble disease Exopthalmus gas bubbles in the gills

Acute bacterial dis. Haemorrhagies Bacteria isolation

PKD Kidney enlargement PKX detection

IHN Haemorrhagies Virus identification

IPN Exopthalmus/ Virus identification

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PREVENTION AND CONTROL

Because no commercial vaccines are available VHS may only be controlled by eradication methods:

COLLECT AND ELIMINATE ALL THE FISH POPULATION DRY ALL THE BASINS SIMULTANEOUSLY (6 WEEKS)
DISINFECT ALL THE FARM
RESTOKE WITH FREE FISH















INFECTIOUS HAEMATOPOIETIC NECROSIS

-IHN -





DEFINITION AND HISTORY

IHN is a systemic viral disease reported, with different synonims, since 1953 and affecting some wild and reared salmonid species along the pacific coast of USA:

YEAR	REGION	DISEASE	AFFECTED SPECIES
1953	Washington	SSVD	O. nerka
1958	Oregon	OSVD	O.nerka
1958	California	SRCD	O. tschawyytscha
1969	Brithish C.	IHN	O. nerka
		IHN	O. mykiss





AETIOLOGY

The causative agent of IHN is a Rhabdovirus, genus Novirhabdovirus 3 main genotypes described (Kurath et al., 2003):

GROUPS ORIGIN

U: Isolates from Alaska, British Columbia Washington,

Oregon, California and Japan obtained from:

Sockeye salmon (O. nerka)

Chinoók salmon (O. tshawýtscha)

M: isolates from Idaho, Washington, France and Italy

obtained from

rainbow trout (O. mikiss)

L: isolates from California, Oregon and Japan obtained from

e Giappone, obtained from :





GEOGRAPHICAL DISTRIBUTION

- CANADA
- USA
- DOMINICAN REP.
- JAPAN
- KOREA
- PAKISTAN
- EUROPE
 - BELGIUM
 - CZECH REPUBLIC
 - GERMANY
 - ITALY
 - FRANCE
 - NETHERLANDS
 - POLAND
 - SLOVENIA





SUSCEPTIBLE HOSTS

According to directive 2006/88 the following species are considered susceptible to the natural disease:

```
rainbow or steelhead trout (O. mykiss),
Chum salmon (Oncorhynchus keta),
coho salmon (O. kisutch),
Masou salmon (O. masou),
sockeye salmon (O. nerka),
pink salmon (O. rhodurus)
Chinook salmon (O.tshawytscha),
Atlantic salmon (Salmo salar
```





TRASMISSION AND PATHOGENESIS

VIRUS ENTRY : <u>Gills</u>, skin, oral

VIRUS SHEDDING : Feces , urine , sessual fluids, mucus

TRASMISSION : Mostly orizontally

Vertical suspected

Confirmed by vectors (invertebrates)

TEMPERATURE: Most of the outbreaks at 8-15° C

REPLICATION: Viremia AT 5-10 days

TARGET ORGANS: haematopoietic tissues (kidney, spleen),

brain and gastro- intestinal.

PROGNOSIS : 35- 40 giorni

MORBIDITY & MORTALITY: 90-95% in fry. Not significant in market-size fish





CLINICAL SIGNS

The first IHN sign is usually represented by a suddenly increase of mortality associated with :

Severe gill anemia

Exopthalmus mono-bilateral

Letargy

Darkening of the skin

Abdomen distension

Mucoid feces

Emorrhagies at the fin base

Deviation of the spinal column in survivors

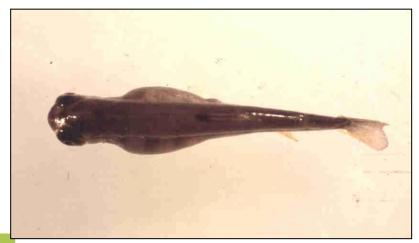












Health and Consumers





Courtesy dr. S. LaPatra

Health and Consumers



GROSS LESIONS

The haemorrhagic picture appearing during an IHN outbreak is usually less severe than in VHS. Affected specimen show:

Visceral anemia

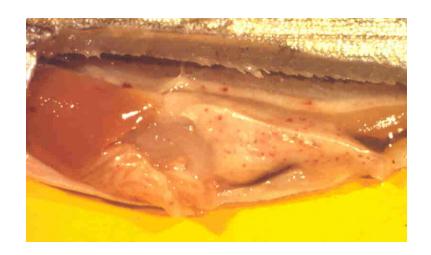
Empty stomach

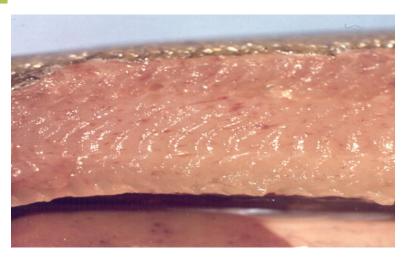
Haemorrhagies in the skin and musculature

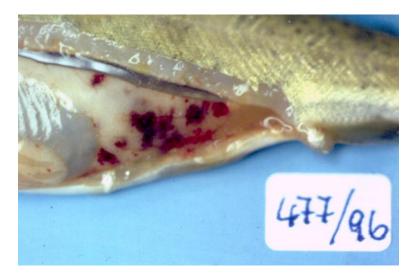
Haemorrhagies in the perivisceral fat















MICROSCOPICAL LESIONS

Necrosis of the haematopoietic tissue Focal necrosis in the liver Necrosis of the pancreatic tissue Necrosis of the granular cells found in the stomach and intestine





DIFFERENTIAL DIAGNOSIS

DISEASE LESIONS TEST

Gas bubble disease Exopthalmus gas bubbles in the gills

Acute bacterial dis. Haemorrhagies Bacteria isolation

PKD Kidney enlargement PKX detection

VHS Haemorrhagies Virus identification

IPN Haemorrhagies Virus identification



PREVENTION AND CONTROL

In addition to the disinfection of eggs , the control of IHN may be obtained by

ERADICATION METHODS

- HARVEST AND ELIMINATE ALL THE FISH POPULATION
- DRY ALL THE BASINS SIMULTANEOUSLY (6 WEEKS)
- DISINFECT
- RESTOKE WITH FREE FISH

VACCINATION

 A DNA VACCINE HAS BEEN REGISTERD IN CANADA TO BE USED IN SALMON INDUSTRY (Salmo salar) .











KOI HERPESVIRUS DISEASE

- KHVD -





HISTORY AND DEFINITION

SINCE 1997-1998 SERIOUS MORTALITIES
AFFECTING KOI CARPS (Cyprinus carpio koi) AND
CHARACTERIZED BY SEVERE NECROSIS OF GILL
EPITELIAL CELLS ASSOCIATED WITH THE
PRESENCE OF HERPESVIRUS PARTICLES HAVE
BEEN OBSERVED

VERY SOON SIMILAR MORTALITIES WERE OBSERVED IN COMMON CARPS (Cyprinus carpio)



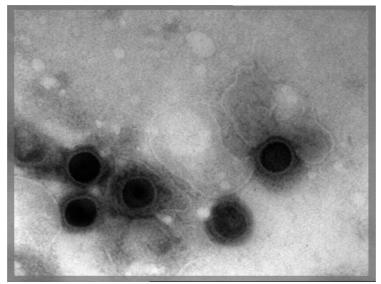


AETIOLOGICAL AGENT

The causative agent of KHVD, is a member of the HERPESVIRIDAE family, identified as:

CyHV-3.

- SUBSPHERICAL MORPHOLOGY
- 150-200 NM DIAMETER
- PRESENCE OF ENVELOPE







SUSCEPTIBLE SPECIES

CYPRINUS CARPIO IS
THE ONLY
SUSCEPTIBLE SPECIES
TO NATURAL KHVD



Common carp (*Cyprinus carpio*)

Common carp (C. carpio koi)

ACCORDING TO RECENT FINDINGS ALL CYPRINID SPECIES SHOULD BE REGARDED AS POTENTIAL CARRIERS OF KHV



Health and Consumers



GEOGRAPHICAL DISTRIBUTION

KHVD HAS BEEN REPORTED FROM THE FOLLOWING COUNTRIES:

Israele	Anonimous, 1998
Germany	Bretzinger et al., 1999
UK	Walster et al., 1999
USA	Hedrick et al., 2000
Belgium	Body et al., 2000
Netherland	2001
China , Malaysia	2001
Denmark	2002
Sumatra; Indonesia , Taiwan	2002
Austria, France, Poland	2003
Luxembourg Switzerland	2003
Japan, Thailand,	2003
South Africa	2003
Singapore	





ORIGIN AND TRASMISSIONE

VIRUS ENTRY : SKIN INCLUDING FINS AND GILLS

VIRUS SHEDDING : SKIN, URINE, FECES,

INCUBATION : 7-21 DAYS

TEMPERATURE : 17-26° C

MORBIDITY: 100%

MORTALITY: \leq 90 %





CLINICAL SIGNS

LETHARGY OR HIPERACTIVITY
SWIMMING CLOSE TO THE WATER SURFACE NEAR TO THE INLET WATER
DISCOLOURATION OR REDDENING OF THE SKIN
FOCAL OR TOTAL LOSS OF THE EPIDERMIS
OVER OR UNDER- PRODUCTION OF MUCUS
SUNKEN EYES
LOSS OF EQUILIBRIUM





European Commission







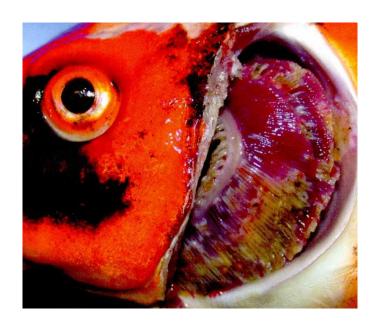


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GROSS AND MICROSCOPIC PATHOLOGY

- SEVERE NECROTIC AREAS IN THE GILLS
- GILLS HYPERPLASIA AND HYPERTROPHY BRANCHIAL EP.
- ADESIONS IN THE ABDOMINAL CAVITY
- ENLARGED KIDNEY AND SPLEEN
- FUSION OF SECONDARY LAMELLAE
- EOSINOPHILIC INCLUSION BODIES



Picture C.E.F.A.S





MORTALITY AND MORBIDITY

MORBIDITY : 100%

MORTALITY : 70-80% IN SEVERE OUTBREAKS 100%

Note: CUMULATIVE MORTALITY MAY BE AFFECTED BY CONCOMITANT BACTERIAL AND PARASITIC INFECTIONS





SAMPLING AND DIAGNOSIS

The diagnosis of KHV must be confirmed by the laboratory

COLLECT 10 FISH WITH TYPICAL KHV SIGNS

POOLED SAMPLES (< 5 FISH EACH) FROM TARGET TISSUES:

KIDNEY

GILLS

PROCESS SAMPLES FOR

VIRUS ISOLATION ON CCB or KF-1 (22-26°C)

PCR (according to Barcovier + Nested)

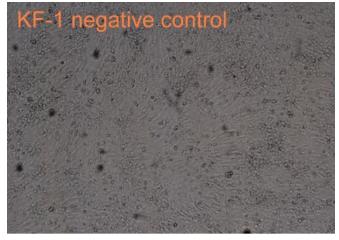
DIFFERENTIAL DIAGNOSIS

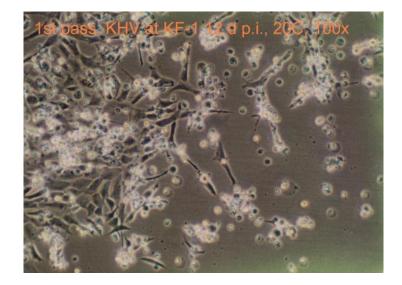
FLAVOBACTERIOSIS

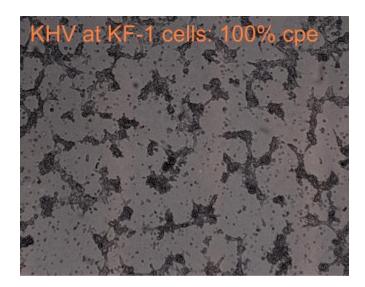




European Commission







By courtesy Dr.Keith Way



CONTROL AND PREVENTION

AN ATTENUATED VACCINE REGISTRED IN ISRAEL

IN ABSENCE OF VACCINE, PREVENTION BY AVOIDING EXPOSURE IS THE ONLY POSSIBILITY

SMALL FARMS MAY ADOPT ERADICATION METHODS





THANK YOU FOR YOUR ATTENTION

giuseppe.bovo@yahoo.it

